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# Malondialdehyde (MDA) Content Assay Kit

**50T/48S**

**Catalogue Number:AK0662-50T-48S**

**Store all reagents at 4°C**

**Validity Period: six months**

**Operation Equipment: Spectrophotometer/Microplate reader**

**For samples:**

**Serum, plasma, cell, Bacteria, fungj, Tissue homogenate.**

**FOR RESEARCH USE ONLY !**

**NOT FOR THERAPEUTIC OR DIAGNOSTIC APPLICATIONS !**

**PLEASE READ THROUGH ENTIRE PROCEDURE BEFORE BEGINNING!**

## **Malondialdehyde (MDA) Content Assay Kit**

**Note: It is necessary to predict 2-3 large difference samples before the formal determination.**

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### **Product Description:**

Lipid peroxide is produced by the action of oxygen free radicals on unsaturated fatty acid, then resolves to compounds, including malondialdehyde (MDA). The level of lipid peroxidation can be showed by detecting the level of MDA.

Under acidic and high temperature conditions, the brown red 3,5,5- three methyl sulfamethoxazole -2,4- two ketone is synthesized with MDA and thiobarbituric acid (TBA) taking place condensation reaction, and the largest absorption wavelength is 532 nm. The content of lipid peroxidation can be estimated after colorimetric. But the soluble sugar will disturb the detection, the production (color reaction of soluble sugar with TBA) have absorption wavelength in 450 nm and 532 nm. In this kit, the MDA content is calculated by the difference between the absorbance at 532 nm, 450 nm and 600 nm.

Because of sucrose in plant tissues and glucose in animal tissues, this kit has two computational formulas for sucrose and glucose. The two formulas are suit for fat.

### **Components:**

**Extraction reagent:** Liquid 60 mL×1. Storage at 4°C.

**Reagent I:** Liquid 42 mL×1. Storage at 4°C.

**Reagent II:** Powder ×2. Storage at 4°C.

**MDA working reagent:** Add 20 mL of Reagent I to Reagent II, dissolve (heat at 70°C or with ultrasonic) and mix thoroughly. Storage at 4°C.

**Reagent III:** Liquid 12 mL×1. Storage at 4°C.

**Note:** The working solution for MDA detection is difficult to dissolve, which can be heated at 70°C and vibrated violently to promote dissolution. Or by ultrasonic treatment to promote dissolution.

### **Reagents and Equipment Required but Not Provided:**

Spectrophotometer, water bath, desk centrifuge, transferpettor, 1 mL glass cuvette, mortar/homogenizer, ice and distilled water.

### **Procedure:**

#### **I. Sample preparation:**

##### **1. Bacteria or cells:**

Collect bacteria or cells into the centrifuge tube. 5 million bacteria or cells could be mixed with 1 mL of Extraction reagent. Use ultrasonication to split bacteria and cells (placed on ice, ultrasonic power 200W,

ultrasonic time 3 seconds, interval 10 seconds, repeat for 30 times). Centrifuge at 8000 ×g for 10 minutes at 4°C to remove insoluble materials, and take supernatant on ice before testing.

2. Tissue:

0.1 g of tissue could be mixed with 1 mL of Extraction reagent and fully homogenized on ice bath. Then centrifuge at 8000 ×g for 10 minutes at 4°C to remove insoluble materials, and take the supernatant on ice before testing.

3. Serum: Detect directly.

**II. Determination procedure:**

1. Preheat spectrophotometer for more than 30 minutes, set zero with distilled water.

2. Add reagents with the following list:

Reagent (μL)	Test tube (T)	Blank tube (B)
MDA working reagent	600	600
Sample	200	-
Distilled water	-	200
Reagent III	200	200

The mixture would be incubated at 100°C for 60 minutes (tightly close to prevent moisture loss), cooled on ice, and centrifuged at 10000 ×g for 10 minutes at room temperature to remove insoluble materials. Take supernatant in 1 mL glass cuvette, and measure the absorbance at 450 nm, 532 nm and 600 nm.  $\Delta A_{450} = A_{450}(T) - A_{450}(B)$ ,  $\Delta A_{532} = A_{532}(T) - A_{532}(B)$ ,  $\Delta A_{600} = A_{600}(T) - A_{600}(B)$ . Blank tube needs to test once or twice.

**III. Calculation:**

1. Tissue, bacteria or cultured cells

1) Protein concentration:

$$\text{MDA (nmol/mg prot)} = (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 1.29 \times \Delta A_{450}) \times V_{rv} \div (C_{pr} \times V_s) \\ = 5 \times (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 1.29 \times \Delta A_{450}) \div C_{pr}$$

2) Sample weight:

$$\text{MDA (nmol/g weight)} = (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 1.29 \times \Delta A_{450}) \times V_{rv} \div (W \times V_s \div V_{sv}) \\ = 5 \times (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 1.29 \times \Delta A_{450}) \div W$$

3) Cell amount:

$$\text{MDA (nmol/10}^4\text{cell)} = (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 1.29 \times \Delta A_{450}) \times V_{rv} \div (400 \times V_s \div V_{sv}) \\ = 0.01 \times (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 1.29 \times \Delta A_{450})$$

4) Serum:

$$\text{MDA (nmol/mL)} = (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 1.29 \times \Delta A_{450}) \times V_{rv} \div V_s \\ = 5 \times (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 1.29 \times \Delta A_{450})$$

2. Plants tissue:

1) Sample weight

$$\text{MDA (nmol/g weight)} = (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 0.56 \times \Delta A_{450}) \times V_{rv} \div (W \times V_s \div V_{sv}) \\ = 5 \times (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 0.56 \times \Delta A_{450}) \div W$$

2) Protein concentration:

$$\text{MDA (nmol/mg prot)} = (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 0.56 \times \Delta A_{450}) \times V_{rv} \div (C_{pr} \times V_s) \\ = 5 \times (6.45 \times (\Delta A_{532} - \Delta A_{600}) - 0.56 \times \Delta A_{450}) \div C_{pr}$$

$V_{rv}$ : Total reaction volume, 1 mL;

$V_s$ : Sample volume, 0.2 mL;

$V_{sv}$ : Extraction volume, 1 mL;

$C_{pr}$ : Sample protein concentration, mg/mL;

$W$ : Sample weight, g;

400: Total number of bacteria and cells, 5 million.

#### **Note:**

If it is found that the absorbance value of the sample is too low, the boiling water bath time can be adjusted from 60 minutes to 90 minutes or longer. The detection of MDA in the same experiment needs to be extended to the same time to avoid errors.

#### **Recent product citations:**

[1] Guoyue Liu, Hong Mei, Miao Chen, et al. Protective effect of agmatine against hyperoxia-induced acute lung injury via regulating lncRNA gadd7. *Biochemical and Biophysical Research Communications*. August 2019; 516(1):68-74.(IF2.705)

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[9] Lijiao Gu, Hantao Wang, Hengling Wei, et al. Identification, Expression, and Functional Analysis

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[10] Ping Shao, Pei Wang, Ben Niu, et al. Environmental stress stability of pectin-stabilized resveratrol liposomes with different degree of esterification. *International Journal of Biological Macromolecules*. November 2018;(IF4.784)

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[1] Spitz D R, Oberley L W. An assay for superoxide dismutase activity in mammalian tissue homogenates[J]. *Analytical Biochemistry*,1989

[2] Masayasu M, Hiroshi Y. A simplified assay method of superoxide dismutase activity for clinical use[J]. *Clinica Chimica Acta*.

